

REVIEW

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# FGFR- gene family alterations in low-grade neuroepithelial tumors



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## Abstract

The discovery of fibroblast growth factor receptor (FGFR) gene family alterations as drivers of primary brain tumors has generated significant excitement, both as potential therapeutic targets as well as defining hallmarks of histologic entities. However, FGFR alterations among neuroepithelial lesions are not restricted to high or low grade, nor to adult vs. pediatric-type tumors. While it may be tempting to consider FGFR-altered tumors as a unified group, this underlying heterogeneity poses diagnostic and interpretive challenges. Therefore, understanding the underlying biology of tumors harboring specific FGFR alterations is critical. In this review, recent evidence for recurrent FGFR alterations in histologically and biologically low-grade neuroepithelial tumors (LGNTs) is examined (namely *FGFR1* tyrosine kinase domain duplication in low grade glioma, *FGFR1-TACC1* fusions in extraventricular neurocytoma [EVN], and *FGFR2-CTNNA3* fusions in polymorphous low-grade neuroepithelial tumor of the young [PLNTY]). Additionally, FGFR alterations with less well-defined prognostic implications are considered (*FGFR3-TACC3* fusions, *FGFR1* hotspot mutations). Finally, a framework for practical interpretation of FGFR alterations in low grade glial/glioneuronal tumors is proposed.

**Keywords:** Low grade glioma, Low grade glioneuronal tumor, FGFR-fusion, FGFR-mutant

## Introduction

The search for disease-defining genetic alterations in brain tumors has characterized the last several decades in neuropathology: one particularly exciting arena has been the discovery of a host of fibroblast growth factor receptor (FGFR) gene family alterations as apparent drivers of primary brain tumors. However, this particular group of lesions has proven especially challenging as they are not confined to either high or low grade, nor to adult vs. pediatric lesions. In fact, FGFR alterations are implicated across a host of human cancers, promoting oncogenesis as a result of overexpression, amplification, mutations, and structural variations [28, 35, 51, 73].

The FGFR family consists of four highly conserved transmembrane tyrosine kinase receptors (*FGFR1–4*) and represents a fundamental receptor tyrosine kinase (RTK) signaling pathway. FGFRs dimerize in the presence of any of 22 known ligands, triggering downstream signaling pathways well-implicated in tumorigenesis; these include the mitogen activated protein kinase

(MAPK) and phosphoinositide-3-kinase (PI3K)/Akt pathways among others [14, 20, 34, 45]. Beyond playing an important role in CNS embryonal development, FGFR signaling influences angiogenesis and tumor cell migration, differentiation, proliferation, and survival. Not surprisingly, FGFRs have emerged as a major target for cancer therapeutics across tumor types and multiple targeting strategies are under investigation [5, 13, 16, 19, 24, 30, 47, 48].

The optimal use of targeted therapy in brain tumors remains under investigation, and its efficacy in low-grade tumors, which would conceivably be slow growing, has been difficult to assess [72]. Although detection of these possible therapeutic targets is of great clinical interest, high quality clinical data remains limited. Ahead of this, understanding the biological implications of specific FGFR alterations, and how this relates to tumor subclassification, is paramount; this is particularly true among histologically low-grade tumors.

Recently the Consortium to Inform Molecular and Practical Approaches to CNS tumor Taxonomy-Not Official WHO (cIMPACT-NOW) released update 4, which specifically addressed so-called “pediatric-type diffuse

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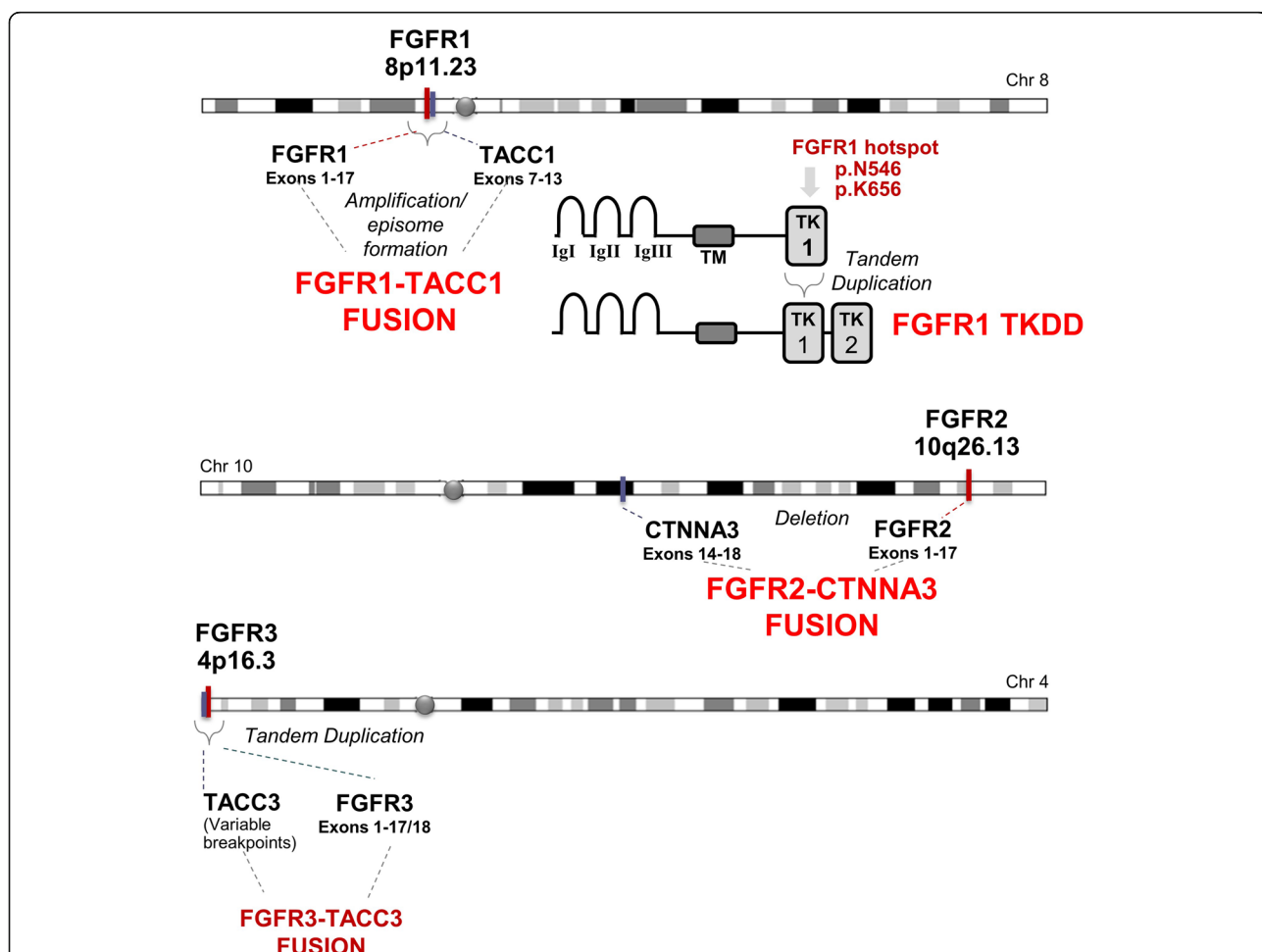
gliomas” [22]. In contrast to the IDH- wild type, diffuse gliomas encountered in adults, diffuse gliomas in children and adolescents most commonly harbor a different constellation of mutations and fusions including alterations in *FGFR1* [56, 77]. The guidelines recommend distinguishing these from adult-type tumors to provide more accurate prognostication, and in some instances guide therapy; delineating relevant diffuse gliomas as harboring either tyrosine kinase domain duplication (TKDD) or single nucleotide variants in *FGFR1*. This is an important step in brain tumor classification and more accurately reflects the relatively prolonged disease course and better overall survival of these pediatric lesions, certainly when compared to IDH-wild type, “adult” tumors. However, while it may be tempting to further consider FGFR-altered tumors as a unified group, there remains significant heterogeneity among them.

In this review, recent evidence for recurrent FGFR alterations in histologically and biologically low grade

neuroepithelial tumors (LGNTs) is examined. These include *FGFR1* tyrosine kinase domain duplication in low grade glioma, *FGFR1-TACC1* fusions in extraventricular neurocytoma (EVN), and *FGFR2-CTNNA3* fusions in polymorphous low grade neuroepithelial tumor of the young (PLNTY). Additionally, FGFR alterations with less well-defined prognostic implications are considered (*FGR3-TACC3* fusions, *FGFR1* hotspot mutations). The structure of these alterations is summarized in Fig. 1. Finally, a proposed framework for interpreting the implications of specific FGFR alterations regarding tumor subclassification and prognostication is presented.

**Genotypic-phenotypic correlations in low grade lesions with FGFR alterations**

Emerging evidence has demonstrated that certain low-grade histologic entities appear to be dominated by specific FGFR alterations. While these mutations have not yet been raised to the level of definitional characteristics by



**Fig. 1** Summary of common FGFR alterations in brain tumors. Some alterations are strongly associated with low grade neuroepithelial lesions: *FGFR1*-TKD, *FGFR1-TACC1* fusion, *FGFR2-CTNNA3* fusion. Others (including *FGFR1* hotspot mutations and *FGFR3-TACC3* fusions) are described in low-grade as well as high-grade tumors, requiring cautious interpretation when encountered in histologic LGNTs

the WHO (and are therefore not required for rendering a diagnosis), there remains (with rare exceptions), a virtual absence in the reported literature of associated high-grade histology and/or aggressive clinical behavior in association with select FGFR alterations. As such, by and large, these alterations may be reasonably regarded as hallmarks of the following low grade neuroepithelial tumors.

#### ***FGFR1- tyrosine kinase domain duplication (FGFR1-TKDD) in low grade glioma (LGG)***

Among the most important insights gained from landmark sequencing studies examining the molecular landscape of pediatric low grade glial and glioneuronal tumors was the identification of an intragenic duplication of the entire *FGFR1* region encoding the tyrosine kinase domain (TKD). This duplication includes exons 10–18 and produces an in-frame fusion separated by a linker element of variable length [56, 77]. Histologically, lesions harboring *FGFR1-TKDD* appear to be predominantly diffuse gliomas located in the cerebral cortex. Duplication of the *FGFR1* TKD has also been reported in low-grade astrocytomas more suggestive of other specific histologic entities including pilocytic astrocytoma (typically extracerebellar) and dysembryoplastic neuroepithelial tumor (DNET, Fig. 2a, b) [23, 37, 40, 60, 77].

While encompassing a significant subset of LGNT (7.4–24%), this alteration appears to be virtually absent in high-grade gliomas (HGG) [38, 77]. In the original report, a cohort of 33 HGG were screened for duplication of the *FGFR1* region encoding the TKD, revealing only one tumor (diagnosed as anaplastic oligoastrocytoma, WHO grade III) that had progressed from a grade II tumor. No *FGFR1-TKDD* positive cases were detected in adult-type oligodendrogliomas, IDH-mutant and 1p/19q co-deleted [77]. Since then, the association of *FGFR1-TKDD* with anaplastic histologic features has proven to be an exceedingly rare phenomenon. One reported case of a rosette forming glioneuronal tumor (RGNT) having focal DNET-like features exhibited multiple local recurrences over a ten-year period, ultimately demonstrated elevated mitoses and high-grade histology, and was shown to harbor *FGFR1-TKDD* in addition to a frameshift mutation in *ATRX* [33]. Additionally, a glioneuronal tumor with features of pilocytic astrocytoma and pleomorphic xanthoastrocytoma also harboring *FGFR1-TKDD* was reported to demonstrate focally elevated mitotic activity; molecular characterization revealed multiple additional variants of unknown significance [3]. It is noteworthy in this instance that, while histologic criteria for anaplasia were met, without long term follow-up data, the biologic and prognostic significance of these findings are unclear. Excepting these rare instances, *FGFR1-TKDD* has been associated with tumors manifesting bland histology and benign clinical behavior.

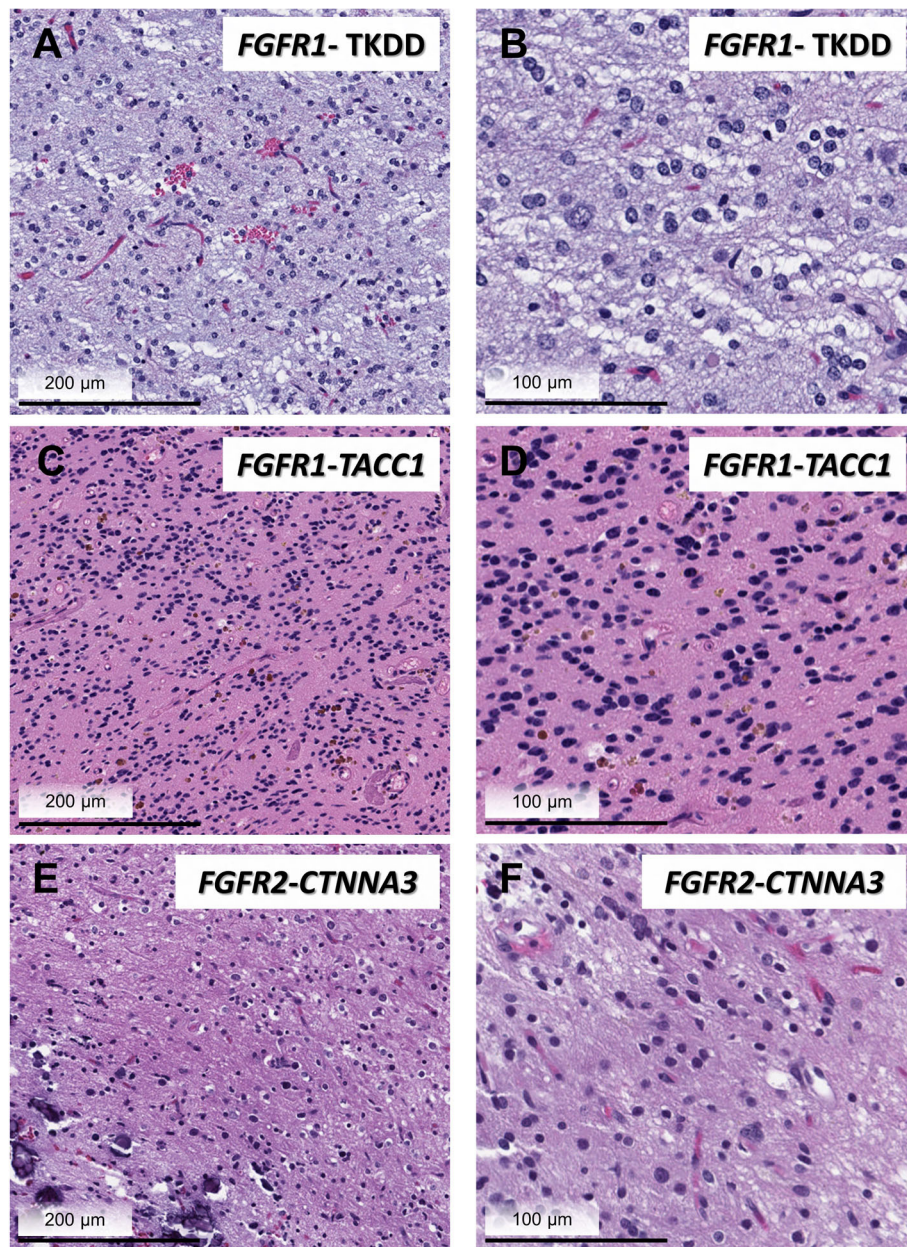
#### ***FGFR1-TACC1 fusion in extra ventricular neurocytoma***

Among the most highly recurrent chromosomal translocations across human cancers are those involving fusions of *FGFR* genes with members of the purportedly oncogenic TACC protein family (*TACC1*, *TACC2*, and *TACC3* [21, 52, 75]). TACC proteins contain a coiled-coil domain at the C-terminus (TACC domain), which facilitates localization of the fusion protein to the centrosome and mitotic spindle [36, 53] in tum promoting aneuploidy and tumorigenesis [49, 69]. Constitutive *FGFR* activity and downstream MAPK/PI3K/mTOR pathway activation also results from the fusion [32, 43].

It is important to note that the highest frequency of *FGFR-TACC* chromosomal translocations is in HGG, namely IDH-wild type GBM where the fusion is between *FGFR3* and *TACC3*, located 48 kb apart on chromosome 4p16 [18, 50, 69] see *FGFR3 fusions*). Among *FGFR*-fusion positive glioblastomas, much less frequently encountered are *FGFR* fusions other than *FGFR3-TACC3*, including *FGFR1-TACC1* [18, 69, 70]. Homologous with regards to respective chromosomal locations, *FGFR1* and *TACC1* are located on chromosome 8p11; the molecular mechanisms with regards to downstream MAPK pathway activation as a result of *FGFR1-TACC1* fusion are also thought to be similar to those of *FGFR3-TACC3*, though less extensively studied and modeled [44].

In sharp contrast to *FGFR3-TACC3*, *FGFR1-TACC1* appears to more commonly associated with low-grade histology and biology, being especially prevalent in the context of extra ventricular neurocytoma (EVN). EVN is a rare primary brain tumor occurring within the parenchyma, outside the ventricular system. While a range of histopathological features may be encountered in EVN, these tumors generally resemble central neurocytoma (Fig. 2c, d). Not surprisingly, accurate diagnosis is confounded by overlapping morphological features with other LGNT entities. DNA methylation-based analysis of a cohort of EVN found that while a subset of histologically diagnosed EVN could be regrouped with other defined, established entities, a large fraction formed a clearly distinct, separate epigenetic group. Importantly, copy number analysis and RNA sequencing demonstrated *FGFR1-TACC1* fusion as a recurrent feature within the EVN methylation group (60%), in addition to a small number of other *FGFR* rearrangements (*FGFR3-TACC3*, *FGFR1-EVI5*) [67].

Indeed, many of the earlier descriptions of EVN predate newer molecular classification of brain tumors and may have been confounded by histologic overlap with other entities. The relationship between rare cases described as *FGFR1-TACC1* fusion-positive HGG/GBM and cases of so-called “atypical EVN” with necrosis, vascular proliferation, and/or elevated mitotic activity, is unclear [25, 29, 41, 44, 69]. The majority of EVNs are



**Fig. 2** Histologic features of FGFR-altered LGNTs. Three examples of LGNTs bearing characteristic FGFR-alteration are shown: DNET with *FGFR1-TKD* (**a, b**), EVN with *FGFR1-TACC1* fusion (**c, d**), and PLNTY with *FGFR2-CTNNA3* fusion (**e, f**). Note that while histologic features of each lesion met diagnostic criteria in keeping with a specific entity, LGNTs share many overlapping histologic features including bland neurocytic/oligodendrogloma-like nuclear features and of lack of significant proliferative or mitotic activity

well-differentiated and generally benign [11]. In the absence of elevated proliferative rate/mitotic activity, and particularly after complete resection, the rate of recurrence is low [25, 41]. While definitive grading criteria have yet to be established and survival data studied in additional independent cohorts, EVN corresponds histologically to WHO grade II, which is in keeping with reported survival data in molecularly-defined EVN, including those bearing *FGFR1-TACC1* fusions [67].

#### *FGFR2- fusion (FGFR2-CTNNA3) in PLNTY*

A recently described entity, “polymorphous low-grade neuroepithelial tumor of the young” or “PLNTY”, has been shown to harbor molecular abnormalities involving the MAPK pathway, including FGFR genes, and a unique fusion involving *FGFR2* [39]. These tumors, while morphologically somewhat variable, are characterized by infiltrative growth, oligodendrogloma-like cytologic features, and frequent calcification (Fig. 2e, f). Strong

cluster of differentiation 34 (CD34) immunohistochemical expression has also been described. Belonging to a group of epilepsy associated low-grade neuroepithelial tumors in children and young adults, PLNTYs appear to have a predilection for the superficial cerebral hemispheres (particularly the temporal lobes), in keeping with prior reports of “long-term epilepsy associated tumors (LEATs)” [10, 39]. Most importantly, all indications point to the indolent behavior of PLNTY [9, 31, 37, 71].

In the original description by Huse et al. (2017) a novel fusion transcript was identified among the series of PLNTY, wherein *FGFR2* (including the kinase domain) joined with exons 14–18 of *CTNNA3* (to include the entirety of its C-terminal dimerization domain) [37, 58]. The oncogenic fusion is thought to result in homodimerization and autophosphorylation of *FGFR2* and downstream *MAPK/PI3K/mTOR* pathway activation, similar to other *FGFR* fusions as previously discussed [15, 69, 71]. Molecular profiling of PLNTYs has demonstrated that they carry a distinct DNA methylation signature, suggesting that they are in fact a distinct biologic entity among at least a subset of LGNTs, including previously described “pediatric oligodendrogliomas” [56, 77]. No reports of *FGFR2-CTNNA3* fusion in association with a high-grade or aggressive tumor have been made to date. However, it is important to note that while *FGFR2-CTNNA3* appears to be relatively specific signature of PLNTY, the molecular landscape of PLNTY includes genetic abnormalities involving either *BRAF* or even *FGFR3*. These other alterations are not unique to PLNTY, and inasmuch as they are frequently also encountered in higher grade entities, should not be regarded as diagnostic of this entity or as predictive of a benign clinical course.

#### Other *FGFR* alterations: unclear implications in LGNT

Several other alterations in *FGFR* genes have been reported in association with LGNTs, but their distribution is not limited to tumors with low grade histology or benign behavior. Therefore, the implications of these alterations in isolation are less clear. Cautious interpretation is advised, particularly in settings where infiltrating or undersampled tumor is a possibility.

#### *FGFR3* fusions

The reality is that the implications of *FGFR3* fusion are clear: as previously stated, *FGFR3* fusions, most commonly *FGFR3-TACC3*, are by and large a feature of IDH-wild type glioblastoma, WHO grade IV [18]. Although *FGFR*-fusion positive GBM constitutes a small subset of GBM as a whole (~ 3%), the sheer preponderance of GBM relative to other types of glioma renders this the most common scenario in which *FGFR3* fusions

will be encountered in most neuropathology practice settings [7, 18, 69].

Difficulty arises when this genetic feature of GBM is encountered in lower grade histologic entities. Detection of *FGFR3* fusions in histologically low-grade tumors is well-documented [18, 27, 37, 38, 77]. However, many of these cases were not been reported with sufficient long-term follow up to determine their clinical biology. This is not to say that *FGFR3* fusions cannot be associated with benign histologic entities; the sole *FGFR3-TACC3* fusion positive case in the original series of PLNTY for example was devoid of any high-grade features suggestive of GBM and demonstrated no evidence of disease or seizures after an extensive interval (89 months) [37]. Of note, *FGFR3-TACC3* fusions in GBM characteristically arise in order individuals, with frequent co-mutation of *TERT* promoter and loss of *CDKN2A/2B*, features that should help distinguish these cases from true LGNT, including PLNTY.

*FGFR3-TACC3* fusion gliomas, both low and high grade, exhibit characteristic histologic features, including monomorphous oligodendroglioma-like nuclei, “chicken-wire” capillary networks, and frequent microcalcifications [7]. While this may be reflective of the common end-result of *FGFR* fusions in all tumors (namely enhanced downstream signaling through *MAPK* kinase pathway effectors), the histologic similarities suggest the possibility of *FGFR3*-fusion positive GBM arising from lower-grade precursor lesions. To date, however, there has been insufficient evidence to support this, and the relationship between high and low grade *FGFR*-fusion positive tumors, if any, remains unclear. Rather, *FGFR3* fusions should prompt a careful evaluation of clinical and neuroradiologic features and call for close surveillance following surgery, when encountered in an ostensibly LGNT.

#### *FGFR1* hotspot (N546 & K656) mutations

Another frequently reported *FGFR* alteration among LGNTs is mutation of two hotspot residues (N546 & K656) in the tyrosine kinase domain of *FGFR1*, well-known to be activating and oncogenic [6, 46, 57, 76]. These two residues are the most commonly mutated residues in *FGFR1* in human cancers and interestingly are described predominately in CNS tumors, mostly histologic pilocytic astrocytomas [40, 78]. Somatic hotspot and germline mutations in *FGFR1* have also been implicated in the pathogenesis of DNET [60]. Of note, encephalocraniocutaneous lipomatosis (ECCL) a sporadic neurocutaneous syndrome with features of disordered *RAS-MAPK* signaling, appears to be mediated in at least a subset of cases by these very *FGFR1* mutations (in mosaic, somatic distribution) and also carries an increased risk of low-grade gliomas, again predominately of

pilocytic astrocytoma histology [6, 8, 42, 54, 64]. It is emerging however, that while these *FGFR1*-mutant tumors certainly can be described histologically and biologically as low grade, they are distinct from typical pilocytic astrocytoma (WHO grade I), which are predominately driven by *BRAF* fusions. In fact, in some early studies, *FGFR1* mutation in pilocytic astrocytoma was associated with a significantly poorer prognosis, although sample size was small [4]. While no specific differentiating histologic criteria have been reported, it has emerged that there are distinguishing clinicopathologic features of these tumors; subsequent larger studies have revealed that pilocytic astrocytoma with *FGFR1* mutation are predominately extracerebellar and frequently midline in location, (in contrast to *BRAF*-fusion positive pilocytic astrocytomas, which predominate in the cerebellum) [40]. At the same time, hotspot *FGFR1* mutations have also been observed in adult and pediatric HGG, at the level of GBM (WHO grade IV) [12, 40, 57]. Notably, *FGFR1* hotspot mutations have been detected in up to 18% of adult midline glioma with high grade histology [55]. These *FGFR1*-mutant HGG frequently demonstrated a recurrent mutational profile in which H3 alterations (H3F3A K27M) and somatic mutations in *NF1* [40] were detected. Although this profile can be seen in tumors histologically equivalent to pilocytic astrocytoma, the underlying molecular features are strongly suggestive of biologic overlap with diffuse midline glioma, H3 K27M-mutant (WHO grade IV) [40, 65].

*FGFR1* hotspot mutations have also emerged as a molecular hallmark of rosette-forming glioneuronal tumor (RGNT) [26, 66]. RGNTs predominately affect young adults and are relatively rare neuroepithelial tumors with distinctive histologic features namely, the presence of neurocytes in rosettes or perivascular pseudo-rosettes in addition to an astrocytic component resembling pilocytic astrocytoma. It is on the basis of histology that the diagnosis is rendered. While in recent studies *FGFR1* hotspot mutations were invariably detected among RGNTs [66], their presence is not currently required for the diagnosis, (and as previously discussed, is certainly not unique to RGNT). Moreover, while RGNT corresponds histologically to WHO grade I and is generally considered benign, dissemination and progression have been reported in rare instances [1, 2, 62, 68, 74]. Of note, frequent co-mutation with *PIK3CA* as well as *NF1* have been reported in RGNT [66]. Mutation of PI3K pathway genes has been associated with aggressive clinical behavior in LGNTs, although further study is needed to determine their prognostic value in RGNT [26, 61]. On the whole, while there is clearly a role for *FGFR1* hotspot mutations in the pathogenesis of LGNT, their specificity for low grade histology and clinical behavior is highly dependent on histologic features and broader molecular context.

### Practical approaches to FGFR alterations in LGNT

Based on available evidence, it appears that some FGFR alterations are more tightly correlated with specific histologic entities among LGNTs, while others may be encountered among variable tumor types, spanning histologic grades and clinical behavior. This poses significant challenges for molecular pathologists, neuropathologists and clinicians: how to determine which amongst these lesions are truly low grade, versus those with increased biologic potential. A practical approach to consider when encountering and “triaging” FGFR alterations in LGNT should involve determining 1) the presence of any atypical features and 2) the presence of additional molecular alterations. Atypical features worth noting in LGNT include both histologic and clinical features. For example, elevated mitotic activity, proliferation indices, and other indicators of high-grade histology should always be noted, even if only focally present in tumors bearing the FGFR alterations described herein. While definitive grading criteria await establishment, in general, bona fide LGNTs are not expected to display significant mitoses, necrosis, or vascular proliferation; proliferative indices would not be expected to exceed 1–2%. Similarly, a multidisciplinary clinical view should be given due consideration in these instances; atypical neuroimaging, and unusual clinical setting (i.e. PLNTY in an older individual [9, 59]) could potentially serve as important indicators of the true nature of the lesion.

By and large, FGFR alterations in LGNTs appear to be a reassuring finding, particularly when they are present in an otherwise genomically quiet background. Most LGNTs appear to be driven by a single molecular pathway, and typically by a single driver genetic alteration [56, 77]. This can be a challenge to definitively determine when taking a minimalist molecular diagnostic approach. Although next-generation sequencing may not be possible to perform in every case, determining the absence of additional alterations (loss of *CDKN2A/2B*, *TERT* promoter mutation, H3- mutation etc.) may be critical to determining the nature of the FGFR-alteration bearing tumor and broader genomic testing should be strongly considered [22].

### Conclusion

While for the purpose of this review, the role of FGFR alterations has been described in relation to specific histologic entities, the reality is that there is significant overlap of histologic features amongst LGNTs (Fig. 2). Although there is utility to the genotypic-phenotypic association between FGFR-alteration and tumor type, it may be more accurate to consider FGFR-altered neuroepithelial lesions as spanning a histologic spectrum. That this group also includes higher grade tumors implies that the spectrum is a biological one as well. Furthermore, it is important to bear

in mind that FGFR- altered tumors are an important subset of a larger group of glial/glioneuronal tumors that are primarily driven by altered MAPK signaling [17, 37, 52, 71].

As previously noted, oncogenic FGFR signaling appears to play a role in a variety of cancer types, including extraneural tumors; FGFR pathway inhibition as a therapeutic strategy remains an area of active investigation. As clinical trials of FGFR inhibitors in brain tumors are ongoing or only recently completed (NCT01975701, NCT028224133, NCT02052778, NCT01948297), we have yet to fully explore the efficacy of this therapeutic approach. Recently, for example, a study found that FGFR inhibitors (AZ4547, dovatinib, PD173074, ponatinib) were more effective in reducing the growth of pediatric diffuse midline glioma, H3 K27M-mutant (diffuse intrinsic pontine glioma, DIPG) cells in vitro compared to Temozolomide [63]. However, much about the role of FGFR inhibitors in treatment of brain tumors, LGNTs in particular, remains to be understood. Optimal design of clinical trials and interpretation of data will be directly dependent on accurate classification of tumors bearing these FGFR alterations.

The complexity of FGFR signaling means that more research will also be necessary to better understand how FGFRs contribute to cancer biology beyond tumor initiation. The role of FGFRs in disease progression as well as associated mechanisms of treatment resistance remain largely unknown (but are certainly relevant issues in the treatment of low grade tumors). With advancing knowledge, we will continue to more accurately identify and stratify LGNTs based on their underlying molecular features, increasingly guiding therapeutic decisions now and in the imminent future.

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#### Competing interests

The author has no competing interests to declare.

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#### References

- Allinson KS, O'Donovan DG, Jena R, Cross JJ, Santarius TS (2015) Rosette-forming glioneuronal tumor with dissemination throughout the ventricular system: a case report. *Clin Neuropathol* 34:64–69
- Anan M, Inoue R, Ishii K, Abe T, Fujiki M, Kobayashi H, Goya T, Nakazato Y (2009) A rosette-forming glioneuronal tumor of the spinal cord: the first case of a rosette-forming glioneuronal tumor originating from the spinal cord. *Hum Pathol* 40:898–901
- Ballester LY, Penas-Prado M, Leeds NE, Huse JT, Fuller GN (2018) FGFR1 tyrosine kinase domain duplication in pilocytic astrocytoma with anaplasia. *Cold Spring Harb Mol Case Stud* 4:a002378
- Becker AP, Scapulatempo-Neto C, Carlioni AC, Paulino A, Sheren J, Aisner DL, Musselwhite E, Clara C, Machado HR, Oliveira RS et al (2015) KIAA1549: BRAF gene fusion and FGFR1 hotspot mutations are prognostic factors in pilocytic astrocytomas. *J Neuropathol Exp Neurol* 74:743–754
- Becker D, Lee PL, Rodeck U, Herlyn M (1992) Inhibition of the fibroblast growth factor receptor 1 (FGFR-1) gene in human melanocytes and malignant melanomas leads to inhibition of proliferation and signs indicative of differentiation. *Oncogene* 7:2303–2313
- Bennett JT, Tan TY, Alcantara D, Tetrault M, Timms AE, Jensen D, Collins S, Nowaczyk MJM, Lindhurst MJ, Christensen KM et al (2016) Mosaic activating mutations in FGFR1 cause encephalocraniocutaneous lipomatosis. *Am J Hum Genet* 98:579–587
- Bielle F, Di Stefano AL, Meyronet D, Picca A, Villa C, Bernier M, Schmitt Y, Giry M, Rousseau A, Figarella-Branger D et al (2018) Diffuse gliomas with FGFR3-TACC3 fusion have characteristic histopathological and molecular features. *Brain Pathol* 28:674–683
- Bieser S, Reis M, Guzman M, Gauvain K, Elbabaa S, Braddock SR, Abdel-Baki MS (2015) Grade II pilocytic astrocytoma in a 3-month-old patient with encephalocraniocutaneous lipomatosis (ECCL): case report and literature review of low grade gliomas in ECCL. *Am J Med Genet A* 167A:878–881. <https://doi.org/10.1002/ajmg.a.37017>
- Bitar M, Danish SF, Rosenblum MK (2018) A newly diagnosed case of polymorphous low-grade neuroepithelial tumor of the young. *Clin Neuropathol* 37:178–181
- Blumcke I, Aronica E, Urbach H, Alexopoulos A, Gonzalez-Martinez JA (2014) A neuropathology-based approach to epilepsy surgery in brain tumors and proposal for a new terminology use for long-term epilepsy-associated brain tumors. *Acta Neuropathol* 128:39–54
- Brat DJ, Scheithauer BW, Eberhart CG, Burger PC (2001) Extraventricular neurocytomas: pathologic features and clinical outcome. *Am J Surg Pathol* 25:1252–1260
- Cancer Genome Atlas Research N (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* 455:1061–1068
- Chae YK, Ranganath K, Hammerman PS, Vakilav C, Mohindra N, Kalyan A, Matsangou M, Costa R, Carneiro B, Villafior VM et al (2017) Inhibition of the fibroblast growth factor receptor (FGFR) pathway: the current landscape and barriers to clinical application. *Oncotarget* 8:16052–16074
- Chen Y, Li X, Eswarakumar VP, Seger R, Lonai P (2000) Fibroblast growth factor (FGF) signaling through PI 3-kinase and Akt/PKB is required for embryoid body differentiation. *Oncogene* 19:3750–3756
- Churi CR, Shroff R, Wang Y, Rashid A, Kang HC, Weatherly J, Zuo M, Zinner R, Hong D, Meric-Bernstam F et al (2014) Mutation profiling in cholangiocarcinoma: prognostic and therapeutic implications. *PLoS One* 9: e115383
- Darwis NDM, Nachankar A, Sasaki Y, Matsui T, Noda SE, Murata K, Tamaki T, Ando K, Okonogi N, Shiba S et al (2019) FGFR signaling as a candidate therapeutic target for cancers resistant to carbon ion radiotherapy. *Int J Mol Sci* 20:4563
- Deng MY, Sill M, Chiang J, Schittenhelm J, Ebinger M, Schuhmann MU, Monoranu CM, Milde T, Wittmann A, Hartmann C et al (2018) Molecularly defined diffuse leptomeningeal glioneuronal tumor (DLGNT) comprises two subgroups with distinct clinical and genetic features. *Acta Neuropathol* 136: 239–253
- Di Stefano AL, Fucci A, Frattini V, Labussiere M, Mokhtari K, Zoppoli P, Marie Y, Bruno A, Boisselier B, Giry M et al (2015) Detection, characterization, and

- inhibition of FGFR-TACC fusions in IDH wild-type glioma. *Clin Cancer Res* 21:3307–3317
19. Dieci MV, Arnedos M, Andre F, Soria JC (2013) Fibroblast growth factor receptor inhibitors as a cancer treatment: from a biologic rationale to medical perspectives. *Cancer Discov* 3:264–279
  20. Doherty P, Walsh FS (1996) CAM-FGF receptor interactions: a model for axonal growth. *Mol Cell Neurosci* 8:99–111
  21. Duncan CG, Killela PJ, Payne CA, Lampson B, Chen WC, Liu J, Solomon D, Waldman T, Towers AJ, Gregory SG et al (2010) Integrated genomic analyses identify *ERRF1* and *TACC3* as glioblastoma-targeted genes. *Oncotarget* 1:265–277
  22. Ellison DW, Hawkins C, Jones DTW, Onar-Thomas A, Pfister SM, Reifenberger G, Louis DN (2019) cIMPACT-NOW update 4: diffuse gliomas characterized by *MYB*, *MYBL1*, or *FGFR1* alterations or *BRAF(V600E)* mutation. *Acta Neuropathol* 137:683–687
  23. Fina F, Baretts D, Colin C, Bouvier C, Padovani L, Nanni-Metellus I, Ouafik L, Scavarda D, Korshunov A, Jones DT et al (2017) Droplet digital PCR is a powerful technique to demonstrate frequent *FGFR1* duplication in dysembryoplastic neuroepithelial tumors. *Oncotarget* 8:2104–2113
  24. Frinchi M, Bonomo A, Trovato-Salinaro A, Condorelli DF, Fuxe K, Spampinato MG, Mudo G (2008) Fibroblast growth factor-2 and its receptor expression in proliferating precursor cells of the subventricular zone in the adult rat brain. *Neurosci Lett* 447:20–25
  25. Furtado A, Arantes M, Silva R, Romao H, Resende M, Honavar M (2010) Comprehensive review of extraventricular neurocytoma with report of two cases, and comparison with central neurocytoma. *Clin Neuropathol* 29:134–140
  26. Gessi M, Moneim YA, Hammes J, Goschzik T, Scholz M, Denkhau D, Waha A, Pietsch T (2014) *FGFR1* mutations in rosette-forming glioneuronal tumors of the fourth ventricle. *J Neuroinflamm Exp Neurol* 73:580–584
  27. Granberg KJ, Annala M, Lehtinen B, Kesseli J, Haapasalo J, Ruusuvoori P, Yli-Harja O, Visakorpi T, Haapasalo H, Nykter M et al (2017) Strong *FGFR3* staining is a marker for *FGFR3* fusions in diffuse gliomas. *Neuro-Oncology* 19:1206–1216
  28. Greulich H, Pollock PM (2011) Targeting mutant fibroblast growth factor receptors in cancer. *Trends Mol Med* 17:283–292
  29. Guerreiro Stucklin AS, Ryall S, Fukuoka K, Zapotocky M, Lassaletta A, Li C, Bridge T, Kim B, Arnoldo A, Kowalski PE et al (2019) Alterations in *ALK/ROS1/NTRK/MET* drive a group of infantile hemispheric gliomas. *Nat Commun* 10:4343
  30. Guillemot F, Zimmer C (2011) From cradle to grave: the multiple roles of fibroblast growth factors in neural development. *Neuron* 71:574–588
  31. Gupta VR, Giller C, Kolhe R, Forseen SE, Sharma S (2019) Polymorphous low-grade neuroepithelial tumor of the young: a case report with genomic findings. *World Neurosurg* 132:347–355
  32. Hadari YR, Gotoh N, Kouhara H, Lax I, Schlessinger J (2001) Critical role for the docking-protein *FRS2* alpha in FGF receptor-mediated signal transduction pathways. *Proc Natl Acad Sci U S A* 98:8578–8583
  33. Halfpenny A, Ferris SP, Grafe M, Woltjer R, Selden N, Nazemi K, Perry A, Solomon DA, Gultekin SH, Moore S et al (2019) A case of recurrent epilepsy-associated rosette-forming glioneuronal tumor with anaplastic transformation in the absence of therapy. *Neuropathology* 39:389–393
  34. Hart KC, Robertson SC, Kanemitsu MY, Meyer AN, Tynan JA, Donoghue DJ (2000) Transformation and stat activation by derivatives of *FGFR1*, *FGFR3*, and *FGFR4*. *Oncogene* 19:3309–3320
  35. Helsten T, Elkin S, Arthur E, Tomson BN, Carter J, Kurzrock R (2016) The *FGFR* landscape in cancer: analysis of 4,853 tumors by next-generation sequencing. *Clin Cancer Res* 22:259–267
  36. Hood FE, Royle SJ (2011) Pulling it together: the mitotic function of *TACC3*. *Bioarchitecture* 1:105–109
  37. Huse JT, Snuderl M, Jones DT, Brathwaite CD, Altman N, Lavi E, Saffery R, Sexton-Oates A, Blumcke I, Capper D et al (2017) Polymorphous low-grade neuroepithelial tumor of the young (PLNTY): an epileptogenic neoplasm with oligodendroglioma-like components, aberrant *CD34* expression, and genetic alterations involving the *MAP* kinase pathway. *Acta Neuropathol* 133:417–429
  38. Johnson A, Severson E, Gay L, Vergilio JA, Elvin J, Suh J, Daniel S, Covert M, Frampton GM, Hsu S et al (2017) Comprehensive genomic profiling of 282 pediatric low- and high-grade gliomas reveals genomic drivers, tumor mutational burden, and hypermutation signatures. *Oncologist* 22:1478–1490
  39. Johnson DR, Giannini C, Jenkins RB, Kim DK, Kaufmann TJ (2019) Plenty of calcification: imaging characterization of polymorphous low-grade neuroepithelial tumor of the young. *Neuroradiology* 61:1327–1332
  40. Jones DT, Hutter B, Jager N, Korshunov A, Kool M, Warnatz HJ, Zichner T, Lambert SR, Ryzhova M, Quang DA et al (2013) Recurrent somatic alterations of *FGFR1* and *NTRK2* in pilocytic astrocytoma. *Nat Genet* 45:927–932
  41. Kane AJ, Sughrue ME, Rutkowski MJ, Aranda D, Mills SA, Lehil M, Fang S, Parsa AT (2012) Atypia predicting prognosis for intracranial extraventricular neurocytomas. *J Neurosurg* 116:349–354
  42. Kocak O, Yazar C, Carman KB (2016) Encephalocraniocutaneous lipomatosis, a rare neurocutaneous disorder: report of additional three cases. *Childs Nerv Syst* 32:559–562
  43. Kouhara H, Hadari YR, Spivak-Kroizman T, Schilling J, Bar-Sagi D, Lax I, Schlessinger J (1997) A lipid-anchored *Grb2*-binding protein that links *FGF*-receptor activation to the *Ras*/*MAPK* signaling pathway. *Cell* 89:693–702
  44. Lasorella A, Sanson M, Iavarone A (2017) *FGFR-TACC* gene fusions in human glioma. *Neuro-Oncology* 19:475–483
  45. LaVallee TM, Prudovsky IA, McMahon GA, Hu X, Maciag T (1998) Activation of the *MAP* kinase pathway by *FGF-1* correlates with cell proliferation induction while activation of the *Src* pathway correlates with migration. *J Cell Biol* 141:1647–1658
  46. Lew ED, Furdul CM, Anderson KS, Schlessinger J (2009) The precise sequence of *FGF* receptor autophosphorylation is kinetically driven and is disrupted by oncogenic mutations. *Sci Signal* 2:ra6
  47. Li M, Bernard O (1992) *FDC-P1* myeloid cells engineered to express fibroblast growth factor receptor 1 proliferate and differentiate in the presence of fibroblast growth factor and heparin. *Proc Natl Acad Sci U S A* 89:3315–3319
  48. Marseglia G, Lodola A, Mor M, Castelli R (2019) Fibroblast growth factor receptor inhibitors: patent review (2015–2019). *Expert Opin Ther Pat* 29:965–977
  49. Nelson KN, Meyer AN, Siari A, Campos AR, Motamedchaboki K, Donoghue DJ (2016) Oncogenic gene fusion *FGFR3-TACC3* is regulated by tyrosine phosphorylation. *Mol Cancer Res* 14:458–469
  50. Parker BC, Annala MJ, Cogdell DE, Granberg KJ, Sun Y, Ji P, Li X, Gumin J, Zheng H, Hu L et al (2013) The tumorigenic *FGFR3-TACC3* gene fusion escapes *miR-99a* regulation in glioblastoma. *J Clin Invest* 123:855–865
  51. Patani H, Bunney TD, Thiyagarajan N, Norman RA, Ogg D, Breed J, Ashford P, Potterton A, Edwards M, Williams SV et al (2016) Landscape of activating cancer mutations in *FGFR* kinases and their differential responses to inhibitors in clinical use. *Oncotarget* 7:24252–24268
  52. Pekmezci M, Villanueva-Meyer JE, Goode B, Van Ziffle J, Onodera C, Grenert JP, Bastian BC, Chamyam G, Maher OM, Khatib Z et al (2018) The genetic landscape of ganglioglioma. *Acta Neuropathol Commun* 6:47
  53. Peset I, Vernos I (2008) The *TACC* proteins: *TACC*-ling microtubule dynamics and centrosome function. *Trends Cell Biol* 18:379–388
  54. Phi JH, Park SH, Chae JH, Wang KC, Cho BK, Kim SK (2010) Papillary glioneuronal tumor present in a patient with encephalocraniocutaneous lipomatosis: case report. *Neurosurgery* 67:E1165–E1169
  55. Picca A, Berzero G, Bielle F, Touat M, Savatovsky J, Polivka M, Trisolini E, Meunier S, Schmitt Y, Idbaih A et al (2018) *FGFR1* actionable mutations, molecular specificities, and outcome of adult midline gliomas. *Neurology* 90:e2086–e2094
  56. Qaddoumi I, Orisme W, Wen J, Santiago T, Gupta K, Dalton JD, Tang B, Hauptfear K, Punchihewa C, Easton J et al (2016) Genetic alterations in uncommon low-grade neuroepithelial tumors: *BRAF*, *FGFR1*, and *MYB* mutations occur at high frequency and align with morphology. *Acta Neuropathol* 131:833–845
  57. Rand V, Huang J, Stockwell T, Ferreira S, Buzko O, Levy S, Busam D, Li K, Edwards JB, Eberhart C et al (2005) Sequence survey of receptor tyrosine kinases reveals mutations in glioblastomas. *Proc Natl Acad Sci U S A* 102:14344–14349
  58. Rangarajan ES, Izard T (2013) Dimer asymmetry defines *alpha*-catenin interactions. *Nat Struct Mol Biol* 20:188–193
  59. Riva G, Cima L, Villanova M, Ghimenton C, Sina S, Riccioni L, Munari G, Fassan M, Giangaspero F, Eccher A (2018) Low-grade neuroepithelial tumor: unusual presentation in an adult without history of seizures. *Neuropathology* 38:557–560
  60. Rivera B, Gayden T, Carrot-Zhang J, Nadaf J, Boshari T, Faury D, Zeinieh M, Blanc R, Burk DL, Fahiminiya S et al (2016) Germline and somatic *FGFR1* abnormalities in dysembryoplastic neuroepithelial tumors. *Acta Neuropathol* 131:847–863
  61. Rodriguez EF, Scheithauer BW, Giannini C, Rynearson A, Cen L, Hoesley B, Gilmer-Flynn H, Sarkaria JN, Jenkins S, Long J et al (2011) *PI3K/AKT* pathway



- alterations are associated with clinically aggressive and histologically anaplastic subsets of pilocytic astrocytoma. *Acta Neuropathol* 121:407–420
62. Schlamann A, von Bueren AO, Hagel C, Zwiener I, Seidel C, Kortmann RD, Muller K (2014) An individual patient data meta-analysis on characteristics and outcome of patients with papillary glioneuronal tumor, rosette glioneuronal tumor with neuropil-like islands and rosette forming glioneuronal tumor of the fourth ventricle. *PLoS One* 9:e101211
  63. Schramm K, Iskar M, Statz B, Jager N, Haag D, Slabicki M, Pfister SM, Zaplatka M, Gronych J, Jones DTW et al (2019) DECIPHER pooled shRNA library screen identifies PP2A and FGFR signaling as potential therapeutic targets for DIPGs. *Neuro-Oncology* 21:867
  64. Schubbert S, Shannon K, Bollag G (2007) Hyperactive Ras in developmental disorders and cancer. *Nat Rev Cancer* 7:295–308
  65. Schwartzentruber J, Korshunov A, Liu XY, Jones DT, Pfaff E, Jacob K, Sturm D, Fontebasso AM, Quang DA, Tonjes M et al (2012) Driver mutations in histone H3.3 and chromatin remodelling genes in paediatric glioblastoma. *Nature* 482:226–231
  66. Sievers P, Appay R, Schrimpf D, Stichel D, Reuss DE, Wefers AK, Reinhardt A, Coras R, Ruf VC, Schmid S et al (2019) Rosette-forming glioneuronal tumors share a distinct DNA methylation profile and mutations in FGFR1, with recurrent co-mutation of PIK3CA and NF1. *Acta Neuropathol* 138:497–504
  67. Sievers P, Stichel D, Schrimpf D, Sahm F, Koelsche C, Reuss DE, Wefers AK, Reinhardt A, Huang K, Ebrahimi A et al (2018) FGFR1:TACC1 fusion is a frequent event in molecularly defined extraventricular neurocytoma. *Acta Neuropathol* 136:293–302
  68. Silveira L, DeWitt J, Thomas A, Tranmer B (2019) Disseminated rosette-forming glioneuronal tumor with spinal drop metastasis, a uniquely aggressive presentation of rare tumor. *World Neurosurg* 132:7–11
  69. Singh D, Chan JM, Zoppoli P, Niola F, Sullivan R, Castano A, Liu EM, Reichel J, Porrati P, Pellegatta S et al (2012) Transforming fusions of FGFR and TACC genes in human glioblastoma. *Science* 337:1231–1235
  70. Stransky N, Cerami E, Schalm S, Kim JL, Lengauer C (2014) The landscape of kinase fusions in cancer. *Nat Commun* 5:4846
  71. Surrey LF, Jain P, Zhang B, Straka J, Zhao X, Harding BN, Resnick AC, Storm PB, Buccoliero AM, Genitori L et al (2019) Genomic analysis of dysembryoplastic neuroepithelial tumor spectrum reveals a diversity of molecular alterations dysregulating the MAPK and PI3K/mTOR pathways. *J Neuropathol Exp Neurol* 78:1100–1111
  72. Tom MC, Cahill DP, Buckner JC, Dietrich J, Parsons MW, Yu JS (2019) Management for different glioma subtypes: are all low-grade gliomas created equal? *Am Soc Clin Oncol Educ Book* 39:133–145
  73. Wu YM, Su F, Kalyana-Sundaram S, Khazanov N, Ateeq B, Cao X, Lonigro RJ, Vats P, Wang R, Lin SF et al (2013) Identification of targetable FGFR gene fusions in diverse cancers. *Cancer Discov* 3:636–647
  74. Xu J, Yang Y, Liu Y, Wei M, Ren J, Chang Y, Huan Y, Yin H, Xue Y (2012) Rosette-forming glioneuronal tumor in the pineal gland and the third ventricle: a case with radiological and clinical implications. *Quant Imaging Med Surg* 2:227–231
  75. Yao R, Natsume Y, Saiki Y, Shioya H, Takeuchi K, Yamori T, Toki H, Aoki I, Saga T, Noda T (2012) Disruption of Tacc3 function leads to in vivo tumor regression. *Oncogene* 31:135–148
  76. Yoon K, Nery S, Rutlin ML, Radtke F, Fishell G, Gaiano N (2004) Fibroblast growth factor receptor signaling promotes radial glial identity and interacts with Notch1 signaling in telencephalic progenitors. *J Neurosci* 24:9497–9506
  77. Zhang J, Wu G, Miller CP, Tatevossian RG, Dalton JD, Tang B, Orisme W, Punchiheva C, Parker M, Qaddoumi I et al (2013) Whole-genome sequencing identifies genetic alterations in pediatric low-grade gliomas. *Nat Genet* 45:602–612
  78. Cosmic (Catalogue of Somatic Mutations in Cancer) (2019) <https://cancer.sanger.ac.uk/cosmic>. Accessed 5 Dec 2019

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